Soft elasticity of RNA gels and negative Poisson ratio

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(Received 11 October 2007; published 19 December 2007)

We propose a model for the elastic properties of RNA gels. The model predicts anomalous elastic properties in the form of a negative Poisson ratio and shape instabilities. The anomalous elasticity is generated by the non-Gaussian force-deformation relation of single-stranded RNA. The effect is greatly magnified by broken rotational symmetry produced by double-stranded sequences and the concomitant soft modes of uniaxial elastomers.

DOI: 10.1103/PhysRevE.76.061910

PACS number(s): 87.14.Gg, 62.20.Dc, 82.35.Lr

I. INTRODUCTION

The study of the viscoelastic properties of networks of flexible, synthetic polymers has for many years been a central topic of polymer science. The classical Flory theory for the elasticity of rubber and of gels treats these systems as networks of nodes linked by highly flexible chains [1-3]. Scaling relations for the viscoelastic moduli that result from this model have been well confirmed [3]. The study of networks of biopolymers has provided a fresh impetus to the field. Gels of semiflexible biopolymers, like F-actin, were shown to obey novel scaling relations [4]. The focus of the present paper is on the elasticity of a different biopolymer system, namely a network or gel of RNA chains. The folding of smaller RNA molecules has been already extensively discussed in the molecular biology literature in the context of ribozymes [5], but extended RNA gels have not received much attention. The genome of single-stranded (SS) RNA viruses, for instance, may form a very promising small scale realization of an RNA gel, as discussed in the Conclusion. It is the purpose of this paper to present a simple model for RNA gels that indicates that such gels should have rather unusual elastic properties that will distinguish them not only from Flory-type gels but also from gels of semiflexible biopolymers like F-actin.

A single-stranded ("SS") RNA chain can be folded first into a "secondary" structure that consists of the pattern of optimal pairing of the bases of the chain [5]. This secondary structure is represented as a planar, branched graph of duplexed double-stranded ("DS") sequences linked by "bubbles" and "stem loops" composed of unpaired bases. A three-dimensional "tertiary" structure is obtained if one also allows complementary pairing between the bases of different bubbles and stem loops of the secondary structure. The model we will study assumes a highly simplified tertiary network topology composed of an array of rigid rods (the DS complementary sequences) that are linked by flexible chains (the SS sequences). Two flexible chains emerge from either of the two ends of each rod (see Fig. 1). Models of this type have in fact been used to describe the folding kinetics of ribozymes [6]. A key ingredient of our model is that these flexible chains are *not* assumed to have the elastic properties of either a Gaussian chain or a semiflexible wormlike chain. Instead, we will examine the elasticity of the network for a general class of interaction potentials. Specifically, we will consider chains that obey the force-extension curve of SS DNA as measured by single-molecule micromechanics.

The particular network of rods and springs that we propose to investigate as a representation of complexed RNA is displayed in Fig. 1. The rods are centered on an altered square lattice. The angle ϕ that an edge of this lattice makes with respect to the vertical, as shown in Fig. 1, parametrizes the overall characteristics of the lattice. If ϕ , which ranges between 0 and $\pi/2$, is equal to $\pi/4$ the lattice is square. At either of the limits of ϕ , the lattice has collapsed onto itself. The edges of the unit cells of the lattice, which do not represent any physical quantity, are represented by grey lines in the figure. The rods are thick solid lines, and the springs are depicted as dashed lines connecting the ends of neighboring rods. As shown in the figure, the length of the rods is 2*l*.

The model belongs to a class of systems, namely uniaxial and biaxial elastomers, that are known to have unusual elastic properties. The internal rotational degrees of freedom produce what is known as "soft" elasticity in the form of large shape changes under applied fields as well as a vanishing of the Poisson ratio [7]. The anomalous elasticity of networks with broken rotational symmetry is an unavoidable and fundamental feature according to a theorem by Golubovic and Lubensky [8]. One of the aims of the present paper is to



FIG. 1. The network of rods and springs that forms the basis of the negative Poisson ratio system. As indicated in the figure, the length of the sides of the underlying altered cubic lattice is a and the length of the rods is 2l.



FIG. 2. Two types of hexagonal networks of hinged rigid rods. The network shown at the top of the figure exhibits a positive Poisson ratio, in that it expands vertically when compressed horizontally. The network at the bottom contracts vertically when compressed horizontally and thus has a negative Poisson ratio.

examine the consequences of this theorem for a concrete model. Specifically, we will explore precisely what physical properties of the interaction potential characterizes anomalous soft elasticity. We will argue in particular that control of the soft elasticity can be achieved by altering the physical properties of the non-Gaussian springs and that this can be achieved in the context of RNA gels.

The proposed model can be viewed as a modified version of a hexagonal lattice of hinged rods. It is actually known in the material science literature that systems that can be represented by models of this type exhibit unusual elasticity. A conventional hexagonal lattice, shown at the top Fig. 2, will expand vertically when compressed horizontally if the rods are rigid and freely hinged. On the other hand, the network at the bottom of the figure-consisting of nonconvex hexagonal units-contracts vertically when compressed horizontally. The network is thus characterized by a *negative* Poisson ratio. The conditions for constructing a network of hinged rods that exhibits a negative Poisson ratio can be elegantly demonstrated by the popular Hoberman sphere [9], which maintains its spherical shape as it expands and collapses [10]. The generic term for a material with a negative Poisson ratio is "auxetic." A negative Poisson ratio is known to be exhibited by iron pyrites [11], self-avoiding, fixed-connectivity membranes [12], monocrystalline zinc [13], carbon nitride [14], polyethylene foams [15], two-dimensional meshlike systems [16], structures composed of rotating rigid units [17] and in network-embedded composites [18]. Biologically, a negative Poisson ratio has been found to be exhibited by both skin [19] and bone [20].

II. POISSON RATIO OF A TWO-DIMENSIONAL NETWORK OF RODS AND SPRINGS

We will start by exploring the properties of the model shown in Fig. 1 as a two-dimensional network. We will represent a distortion of the lattice in terms of a strain tensor $\vec{\epsilon}$, so that the displacement $\Delta \vec{r}_i = (\Delta x_i, \Delta y_i)$, of a lattice vertex originally at the location $\vec{r}_i = (x_i, y_i)$ is given by



FIG. 3. A triad of neighboring rods, connected in this figure by two springs connected to the top end of the lower central rod. The rods and the lattice are shown in an as-yet undistorted state. The lattice is compressed horizontally as compared to the one shown in Fig. 1.

$$\Delta x_i = \epsilon_{xx} x_i + \epsilon_{xy} y_i \tag{2.1}$$

and similarly for Δy_i . We will also allow the rods to rotate in the plane by the angle θ .

Consider, now, the triad of rods with connecting springs shown in Fig. 3. Note that the underlying lattice is more compressed along the horizontal direction than the lattice shown in Fig. 1. As will be demonstrated below, the degree of alteration of the square lattice is controlled by the requirement that the rod and spring network can be stabilized by an *osmotic* pressure. The distance between the upper end of the lower rod and the lower end of the rod on the right is equal to

$$d_1^2 = (a\epsilon_{xy}\cos\phi - 2l\sin\theta + a\sin\phi + a\epsilon_{xx}\sin\phi)^2 + (-2l\cos\theta + a\cos\phi + a\epsilon_{yy}\cos\phi + a\epsilon_{yx}\sin\phi)^2$$
(2.2)

and the distance between the upper end of the lower rod and the lower end of the rod to the left is given by

$$d_2^2 = (a\epsilon_{xy}\cos\phi - 2l\sin\theta - a\sin\phi - a\epsilon_{xx}\sin\phi)^2 + (-2l\cos\theta + a\cos\phi + a\epsilon_{yy}\cos\phi - a\epsilon_{yx}\sin\phi)^2.$$
(2.3)

The angle θ in Eqs. (2.3) is the angle to which all the rods rotate under the influence of the uniform distortion of the lattice as parametrized by the constant strain tensor $\vec{\epsilon}$.

A. Non-Gaussian behavior of the springs

In order to facilitate numerical calculations, we will parametrize the interactions mediated by the springs in terms of the square of the distance between the spring end points. That is, we express the energy of the interaction in terms of the variable x, where

$$x = d^2 - d_0^2. (2.4)$$

The quantity *d* is the distance between end points, and d_0 is the distance between end points in the equilibrium state. Next, let V(d) be the interaction potential that represents the free energy of an SS sequence connecting two points a distance *d* apart. We will expand V(d) in a Taylor series around the equilibrium state x=0:

$$V(\sqrt{d_0^2 + x}) = V(d_0) + \frac{x}{2d_0}V'(d_0) + \frac{x^2}{8d_0^3}[-V'(d_0) + d_0V''(d_0)] + \cdots$$
$$= \nu_0 + \nu_1 x + \frac{\nu_2}{2}x^2 + \cdots .$$
(2.5)

It is important to note that even if the interaction V(d) is convex upward, so that V''(d) > 0, the second order coefficient v_2 in the expansion above can be negative. Consider, for instance, the case of a simple power-law interaction energy of the form

$$V(d) = Cd^p \tag{2.6}$$

with C a positive coefficient and p a power greater than 1. Then,

$$V'(d_0) = Cpd_0^{p-1} \tag{2.7}$$

and

$$d_0 V''(d_0) - V'(d_0) = Cp(p-2)d_0^{p-1}.$$
 (2.8)

If p < 2, then, assuming that the coefficient *C* is greater than zero, the second order coefficient in Eq. (2.5), ν_2 , will be negative. In the case of Gaussian chains with Hooke-law-type harmonic interaction, ν_2 is equal to *zero*, as are all higher order coefficients in the expansion of the interaction in terms of the variable *x*. On the other hand, for a freely jointed chain or a wormlike chain, $\nu_2 > 0$.

Under the assumption that the interaction mediated by the springs is always attractive, the angle θ at which the rods tilt must adjust in such a way as to minimize the sum of the two distances d_1 and d_2 . Taking the derivative of d_1+d_2 with respect to θ and constructing an extremum equation, we end up with the relationship

$$8al\cos\phi[(\epsilon_{yy}+1)\sin\theta - \epsilon_{xy}\cos\theta] = 0.$$
(2.9)

The strain tensor is assumed to be a small quantity. This means that the solution to the equation above is, to an accuracy sufficient for our purposes,

The total energy associated with the interactions mediated by the springs shown in Fig. 3 is then given by

$$E = \nu_0 + \nu_1 [(d_1^2 - d_0^2) + (d_2^2 - d_0^2)] + \frac{\nu_2}{2} [(d_1^2 - d_0^2)^2 + (d_2^2 - d_0^2)^2]$$
(2.11)

with θ in Eqs. (2.2) and (2.3) as given by Eq. (2.10).

B. Determination of the angle ϕ , expansion of the energy in strain coordinates, and calculation of the Poisson ratio

The next step is to expand the resulting expression to second order in the strain tensor. At zeroth order, we are left with the coefficient v_0 . To first order in $\vec{\epsilon}$ we have the following contribution to the energy:

$$4a\nu_1[a\epsilon_{xx}\sin^2\phi + \epsilon_{yy}\cos\phi(a\cos\phi - 2l)]. \quad (2.12)$$

The appearance of terms linear in the strain energy indicates that the system has a tendency to spontaneously deform. First, consider the case of a uniform contraction. In terms of the strain tensor, this yields an energy going as $\epsilon_{xx} + \epsilon_{yy}$. Such a uniform deformation can be countered by an osmotic pressure II that increases as the monomer concentration grows under contraction. If the coefficients of ϵ_{xx} and ϵ_{yy} in Eq. (2.12) are equal to each other then osmotic pressure suffices to completely balance the first order energy for $d=d_0$ (the equilibrium state) in Eq. (2.12). On the other hand, if the coefficients of ϵ_{xx} and ϵ_{yy} are unequal, then the system undergoes a spontaneous shear transformation, which can only be balanced by the application of anisotropic stress.

We assume that the only external stress acting on the network is due to osmotic pressure. Given this presumption, the resulting requirement on the linear coefficients in Eq. (2.12) translates into an equation for the lattice angle ϕ that is satisfied when

$$\phi = \arccos\left[\frac{l}{2a} + \sqrt{\left(\frac{l}{2a}\right)^2 + \frac{1}{2}}\right].$$
 (2.13)

Two limits of the above expression are noteworthy. When l = 0, so the rods are infinitesimal in extent, then ϕ as given by Eq. (2.13) is equal to $\pi/4$, consistent with an underlying square lattice. On the other hand, when l=a/2, we find a ϕ from Eq. (2.13) that is equal to zero, which implies a horizontal collapse of the complex. In this limit, the rods are exactly long enough that their tips touch in the event of such a collapse of the lattice. Henceforth we will assume that l lies in the range between 0 and a/2.

Assuming cancellation of the linear term, the energy is now, at lowest nontrivial order, quadratic in the elements of $\vec{\epsilon}$. This contribution to the quadratic energy is of the form

$$\Omega[\boldsymbol{\epsilon}] = \sum_{i,j,k,l} \alpha_{ij,kl} \boldsymbol{\epsilon}_{ij} \boldsymbol{\epsilon}_{kl}.$$
 (2.14)

In light of translational invariance, this energy will be the same for the energy supplied by the two springs attached to the upper tip of every rod in the complex, under the assumption of a uniform strain. We have thus effectively calculated the energy of interaction in the entire complex.



FIG. 4. The two Poisson ratio of the two-dimensional lattice according to Eq. (2.17).

The strain induced by an externally applied stress $\vec{\sigma}$ is the solution to the set of equations

$$\frac{\partial}{\partial \epsilon_{ij}} \Omega[\vec{\epsilon}] = \sigma_{ij}. \tag{2.15}$$

We will take the strain to be exerted in the y direction, so σ_{yy} is the only nonzero element of the strain tensor. After solving Eq. (2.15) for the elements of the strain tensor we can extract the Poisson ratio μ via the relationship

$$\mu = -\frac{\epsilon_{xx}}{\epsilon_{yy}}.$$
 (2.16)

Following a straightforward calculation we obtain the explicit result

$$\mu = \frac{a^2(a^2 - 4l^2)\nu_2}{[a^2 - l^2 + al\sqrt{2} + (l/a)^2]\nu_1 + a^2(a^2 - 4l^2)\nu_2}.$$
(2.17)

C. Poisson ratio when there are no rods

It is useful to consider certain limiting cases of Eq. (2.17). First, consider the case l=0, which corresponds to a network in which the rods are replaced by point contacts. If we set l=0 in the expression Eq. (2.17), the expression for the Poisson ratio further simplifies to

$$\frac{a^2 \nu_2 / \nu_1}{1 + a^2 \nu_2 / \nu_1} \tag{2.18}$$

which predicts a negative Poisson ratio, if the ratio ν_2/ν_1 is negative. To check for stability of the lattice one can determine the eigenvalues of the energy matrix \vec{a} . When l=0, this four-by-four matrix is degenerate, and there are two distinct eigenvalues: $a^2\nu_1 - a^4\nu_2$ and $a^2\nu_1 + 3a^4\nu_2$. Both eigenvalues are positive, corresponding to a stable lattice, provided $-\nu_1/3a^2 < \nu_2 < \nu_1/a^2$, where we assume that $\nu_1 > 0$. For ν_2 in this range, the Poisson ratio varies from -1/2 to 1/2. It follows that a moderately negative Poisson ratio is, in fact, possible in a lattice consisting entirely of non-Gaussian springs. Note that this is not related to the existence of internal rotational degrees of freedom.



FIG. 5. The determinant of the elastic energy matrix $\vec{\alpha}$ when l = 0.45a, and ν_1 and a have been set equal to 1.

D. Poisson ratio at general *l*

We now set l=0.45a. This leads to an equilibrium state that is compressed horizontally as in Fig. 3. In Fig. 4 we show μ as a function of ν_2 .

We have again checked the stability by computing the determinant of the four-by-four matrix with elements $\alpha_{ij,kl}$. Figure 5 shows a typical plot of the determinant as a function of ν_2 . The determinant vanishes at ν_2 =-6.232, corresponding to the emergence of a negative eigenvalue of the energy matrix $\vec{\alpha}$ and the development of a mechanical instability in the complex [21]. At that threshold value of ν_2 , we find μ =-4.212 26, so a substantial negative Poisson ratio is indeed possible close to a mechanical instability. Figure 6 shows the determinant for an extended range of the interaction coefficient ν_2 . As illustrated in that figure, the range of stability of the energy matrix is bounded from above as well as below as a function of ν_2 .

If we compare these results with those in the case l=0 we conclude that the broken rotational symmetry for $l \neq 0$ has greatly *amplified* the negative Poisson ratio produced by a negative value of ν_2 . There is indeed no intrinsic physical bound on the Poisson ratio in an anisotropic solid [22]. The amplification effect is most dramatic when ν_2 approaches the threshold of the mechanical instability, at $\nu_2=-6.232$. Note though that according to Eq. (2.17) the Poisson ratio *formally* is zero even at this critical point for a Gaussian network and any other system with $\nu_2=0$. It should be recalled here that the Poisson ratio is zero in conventional nematic elastomers [7]. One can compare the effect of a negative ν_2 to that of a



FIG. 6. The determinant of the energy matrix $\vec{\alpha}$ as a function of ν_2 , with l=0.45a and a and ν_1 set equal to 1. The range of stability is indicated by the heavy line on the horizontal axis.



FIG. 7. Portion of the cubic lattice, shown unsheared here, with the rods at the vertices. The "springs" connecting the vertices are shown as dashed lines. The edges of the cubic lattice in which the rods are embedded are shown as grey lines. Note that these edges do not represent actual physical structures. Only those springs that connect two rods shown in the figure are depicted. Finally, a principle diagonal, lying along the z axis, is shown as a dashed and dotted line.

small magnetic field applied to a magnet as one approaches the Curie temperature. The divergence of the susceptibility amplifies the effect of the applied field. In the next section we will extend the notion of the sort of complex we have been discussing to a three-dimensional system.

III. THREE-DIMENSIONAL NETWORK OF RODS AND SPRINGS EMBODYING A NEGATIVE POISSON RATIO

A. Preliminaries: Description of parameters

The way in which the three-dimensional network is constructed will be a generalization of the two-dimensional case. Imagine a sheared version of a cubic lattice rotated so that the z-axis is along a diagonal. A rod is placed at each vertex of this lattice, and each end of this rod is connected to the ends of the three rods that are closest to it by a spring having a nonlinear force-extension relation. Figure 7 depicts a portion of this lattice of rods, shown as short solid lines. The springs are represented by dashed lines and the edges of the lattice-shown for illustrative purposes only-are represented by grey lines. The z axis is along the principal diagonal, shown as a grey dashed and dotted line in the figure, and the x axis is aligned with the projection in the x-y plane of an edge of the cube. Three neighboring vertices of the cube are indexed as shown in Fig. 8. If one were to interpret the configuration illustrated in Figs. 7 and 8 in terms of an RNA network, then, given that each rod is attached to three strands, it would be necessary to assume that the rods represent "triplexed" segments of the molecule. Discarding this exotic version of complexed RNA, we interpret the organization illustrated there as a kind of "averaged" version of a structure in which each rod terminates in two springs, as shown in Fig. 1. The actual connectivity is approximated by the three springs shown in Fig. 8, each carrying a fraction of



FIG. 8. Three vertices of the cubic lattice with associated rods and springs attached to them. The numbers are the indices of the vertices. As in Fig 7, the springs are shown as dashed lines.

the restoring energy stored in the two springs in the actual structure.

The rod with index 0 is assumed to be located at the origin. The coordinates of the corners on which the neighboring rods sit are given by

$$x_1 = a \cos \psi, \tag{3.1}$$

$$y_1 = 0,$$
 (3.2)

$$z_1 = a \sin \psi, \tag{3.3}$$

$$x_2 = A \cos \psi \cos(2\pi/3),$$
 (3.4)

$$y_2 = a \cos \psi \sin(2\pi/3),$$
 (3.5)

 $z_2 = a \sin \psi, \tag{3.6}$

$$x_3 = a \cos \psi \cos(4\pi/3),$$
 (3.7)

$$y_3 = a \cos \psi \sin(4\pi/3),$$
 (3.8)

$$z_3 = a \sin \psi. \tag{3.9}$$

The quantity *a* in Eqs. (3.1)–(3.9) is the length of a cube edge. The quantity ψ is the angle that the edges of the cube in Fig. 8 make with respect to the *x*-*y* plane. In the case of an unsheared cubic lattice, this angle is equal to $\arctan(1/\sqrt{2})$. Note that when $\psi = \pi/2$, the edges connecting the site at the origin to the three neighboring ones are all vertical, so that the neighboring sites all lie at exactly the same location. Each edge in the cubic lattice has length *a* and the length of the rods is 2*l*, again as shown in Fig. 8. Initially, the rods point along the *z* axis. However, we will assume that when the lattice shears the rods change orientation. The direction in which the rods point will be described in terms of the standard spherical angles θ and θ . The next step is to allow the lattice to distort. As in the previous section we represent the



FIG. 9. The angles ϕ and θ . All three edges shown in the figure have an orientation with respect to the horizontal plane.

distortion in terms of a strain tensor $\vec{\epsilon}$, so that placing one of the vertices of the lattice at the origin, the displacement $\Delta \vec{r_i} = (\Delta x_i, \Delta y_i, \Delta z_i)$, of any other lattice vertex originally at the location $\vec{r_i} = (x_i, y_i, z_i)$ is given by

$$\Delta x_i = \epsilon_{xx} x_i + \epsilon_{xy} y_i + \epsilon_{xz} z_i \tag{3.10}$$

and similarly for Δy_i and Δz_i .

B. Determination of the angles ψ , ϕ , and θ

First, we calculate the quantities (x_i, y_i, z_i) for each of the springs shown in Fig. 8. Then, we expand the energy to first and second order in the (x_i, y_i, z_i) . The resulting expression is then expanded once again, now to second order in both the strain tensor $\vec{\epsilon}$ and the rotation angle θ . As both quantities are expected to be small, we restrict our consideration to terms that are at most quadratic in either one or both of them. The next step is to determine the angle ψ , (shown in Fig. 9) by which the lattice shears. This angle is again controlled by the requirement that the lattice, in the absence of any imposed stress, can be stabilized by an osmotic pressure Π , that couples only to the total volume of the lattice, or in other words to the combination Tr $\vec{\epsilon} = \epsilon_{xx} + \epsilon_{yy} + \epsilon_{zz}$.

If we expand the energy of the lattice to first order in $\hat{\epsilon}$ and zeroth order in θ , we obtain the following expression:

$$3a^{2}(\epsilon_{xx} + \epsilon_{yy})\cos^{2}\psi + 6a\epsilon_{zz}\sin\psi(a\sin\psi - 2l).$$
(3.11)

The requirement that this reduce to a function of the trace of the strain tensor only leads to a quadratic equation for $\sin \psi$. The solution to this equation is

$$\psi = \arcsin\left[\frac{2l + \sqrt{3a^2 + 4l^2}}{3a}\right] = \Psi_0(l/a).$$
 (3.12)

It is useful to consider the two limits of the above expression. When $l \rightarrow 0$, the argument of the arcsin reduces to $1/\sqrt{3}$, which is consistent with an arctangent equal to $1/\sqrt{2}$ and a cubic lattice. On the other hand, as $l \rightarrow a/2$, the angle ψ goes to $\pi/2$, which means that the lattice collapses onto itself. In the latter limit, the rods are long enough that they touch end to end when that collapse takes place.

Inserting the value of ψ given by the right hand side of Eq. (3.12) into the expression (3.11), we find for the dependence of the energy on uniform dilations of the lattice

$$\frac{2}{3}(\epsilon_{xx} + \epsilon_{yy} + \epsilon_{zz})[3a^2 - 4l^2 - 2l\sqrt{3a^2 + 4l^2}] \quad (3.13)$$

so the equation of state of the system is

$$\Pi v_0 = \frac{2}{3} \nu_1 [3a^2 - 4l^2 - 2l\sqrt{3a^2 + 4l^2}], \qquad (3.14)$$

$$v_0 = a^3 (3\sqrt{3}/2) \cos^2 \psi \sin \psi = \frac{(3a^2 - 8l^2)\sqrt{3a^2 + 4l^2} - 16l^3}{3\sqrt{3}}$$
(3.15)

is the volume per rod in the lattice. As $l \rightarrow a/2$, the expression multiplying the trace of the strain tensor vanishes, and the linear dependence of the energy on uniform dilations of the lattice is lost. The osmotic pressure Π approaches the limiting value $2\nu_1/\sqrt{3}a$.

In light of this initial adjustment of the lattice, the next step is to determine the extent to which the lattice responds to an external stress. The response manifests itself in three quantities: the strain tensor $\vec{\epsilon}$, the rod tilt angle θ , and the azimuthal angle ϕ , of the rods. As it turns out, the azimuthal angle appears only in the term that is first order in θ , and the form of that term is $A \cos \phi + B \sin \phi$, where A and B are linear functions of the strain tensor and general functions of the angle ψ . This expression is minimized when ϕ = arctan A/B and is equal in this case to $-\sqrt{A^2 + B^2}$. The second-order-in- θ contribution to the energy of the deformed lattice is equal to

$$6al\{2al\nu_2[\cos\Psi_0(l/a)]^2 + \nu_1\sin\Psi_0(l/a)\}\theta^2 \equiv D_2\theta^2.$$
(3.16)

If we denote by D_0 the portion of the deformation energy that is zeroth order in θ and second order in the strain tensor, the net free energy as a function of the polar angle is

$$D_0 - \theta \sqrt{A^2 + B^2} + D_2 \theta^2.$$
 (3.17)

Minimizing Eq. (3.17) with respect to θ , we end up with our final result for the dependence of the energy on the strain tensor $\vec{\epsilon}$ in the uniformly strained lattice. In terms of the quantities defined above, the energy has the form

$$D_0 - \frac{A^2 + B^2}{4D_2} = \sum_{i,j,k,l} \alpha_{ij,kl} \epsilon_{ij} \epsilon_{kl} \equiv \Omega[\vec{\epsilon}].$$
(3.18)

As indicated in Eq. (3.18), the energy of the uniformly distorted lattice with the angles ψ , ϕ , and θ relaxed to the values dictated by energy minimization and lattice stabilization is purely quadratic in the strain tensor. The tensorial quantity $\alpha_{ij,kl}$ is symmetric with respect to interchange of the index pairs *ij* and *kl*. The response of the system of rods and springs to an externally generated stress tensor $\vec{\sigma}$ follows from the solution to the set of linear equations

$$\frac{\partial \Omega[\vec{\epsilon}]}{\partial \epsilon_{ij}} = \sigma_{ij}.$$
(3.19)

Given the overall anisotropy of the lattice, symmetry arguments yield only restricted information. If we orient the *z* axis along the direction defined by the rods in the unstressed lattice, then symmetry arguments tell us that a stress entirely in the *z* direction results in equal values of the strain tensor components ϵ_{xx} and ϵ_{yy} , while $\epsilon_{xy} = \epsilon_{yx} = 0$.

C. Poisson ratios for l=0

Once again we consider the case of a lattice consisting of springs only, in which the rods are replaced by point connec-

where



FIG. 10. The Poisson ratio as a function of $a^2\nu_2/\nu_1$ when l=0. The asymptote of 1 is also indicated.

tions. For l=0, the Poisson ratio of interest becomes

$$\mu = \frac{4(a^2\nu_2/\nu_1)}{3 + 4(a^2\nu_2/\nu_1)} \tag{3.20}$$

and the determinant of the energy matrix is

det
$$\vec{\alpha} = 512a^{18}\nu_1^9 \left(1 + \frac{a^2\nu_2}{\nu_1}\right)^3$$
. (3.21)

This means the energy matrix is associated with a stable equilibrium state as long as $a^2\nu_2/\nu_1 > -1/2$. Figure 10 is a plot of the Poisson ratio as given by Eq. (3.20) in the range of stability of the energy. The limits of this quantity are 1 as $a^2\nu_2/\nu_1 \rightarrow \infty$ and -2 as $a^2\nu_2/\nu_1 \rightarrow -1/2$, which are the limits of the Poisson ratio in an isotropic solid. This means that, as in two dimensions, broken rotational symmetry is not essential for the negative Poisson ratio.

D. Poisson ratios for $l \neq 0$

To explore the properties of the Poisson ratio for $l \neq 0$, we will focus on the exponent *p* in the power law interaction (2.6). Consider the graph shown in Fig. 11, which is of the Poisson ratio, defined as

$$\mu = -\frac{\epsilon_{xx} + \epsilon_{yy}}{\epsilon_{zz}} \tag{3.22}$$

in the case of a stress entirely in the *z* direction ($\sigma_{ij}=0$ except for σ_{zz}). The length *l* of the rod has been set equal to 0.4*a*. Note that the Poisson ratio passes through zero as the power law passes through the power associated with the Gaussian spring. When p < 2, the Poisson ratio is negative, which means that the lattice system resists shearing.



FIG. 11. Graph of the Poisson ratio vs the exponent p of the power law in Eq. (2.6).



FIG. 12. The Poisson ratio, plotted against the exponent p in Eq. (2.6). Here p ranges from 0 to 3. As noted in the text, a value of p less than 1 is inconsistent with mechanical stability.

Figure 12 is a graph of the Poisson ratio when l=0.45a as a function of p. The range of stability, i.e., of a positive determinant for $\vec{\alpha}$, is for p > 1, with p=1 the location of the onset of instability. Recall that in the two-dimensional case, μ approaches a finite negative value at the point of instability. From Fig. 12 we see that the Poisson ratio *diverges to* $-\infty$ *at the instability threshold.* The amplification effect of the rotational degree of freedom in producing a large negative Poisson ratio is thus much more pronounced in three dimensions. By "tuning" the physical properties of the non-Gaussian springs it is possible to dramatically alter the elastic response of the network.

IV. RNA IN NETWORKS AND A NEGATIVE POISSON RATIO

In this conclusion, we will apply the results of the last section to RNA networks and discuss the consequences in terms of soft elasticity and negative Poisson ratios. In Secs. II and III we learned that ν_2 functions as a control parameter for elasticity. If ν_2 is positive, one obtains conventional elasticity. As noted in the Introduction, gels of flexible and semiflexible polymers are conventionally modeled as networks of Gaussian chains, wormlike chains or freely jointed chains, all of which have $\nu_2 > 0$. If ν_2 is negative, we predict a range of anomalous elasticity, with negative Poisson ratios terminating in a mechanical instability at a critical value for ν_2 , at which point μ diverges to $-\infty$. Is it realistic for a biopolymer network to have a negative ν_2 ? The ν_2 parameter can in principle be determined from the force-extension curves of the nonlinear springs. For an RNA network of the type shown in Fig. 1, this would be the force-extension curve of single stranded RNA. Force-extension curves have been measured for single-stranded RNA and folded RNA molecules [23] but not yet for long single-stranded chains. Such measurements have been performed for single-stranded DNA strands of about 10^4 bases [24]. A typical set of results, taken from [24] is shown in Fig. 13. The force-extension curve rises rapidly when the extension reaches the contour length. This is preceded by a range of extensions for which the force is relatively constant, which may be due to progressive loss of stacking interaction. The force-extension curve for small extensions is presumably dominated by entropic elasticity. We fitted the measured data with a fifth order polynomial



FIG. 13. Force vs extension data for a segment of singlestranded charomid DNA [25] with a backbone length of 5.7 μ m in one millimolar phosphate buffer. The data were extracted from Fig. 1 of [24]. Also shown in this figure is a fifth order polynomial fit to the data.

form, from which we were able to extract the quantities ν_1 (in pN/ μ m²) and ν_2 (in pN/ μ m⁴), shown in Figs. 14 and 15.

The ν_2 control parameter is indeed negative for extensions less than 3 μ m. It should be noted that ν_1 and ν_2 in general are expected to be quite sensitive to solvent conditions. We can now make use of the methods described above to compute the Poisson ratio for given ratio l/a, and we find that there is indeed a negative Poisson ratio over the full range of values of the length parameter *a*, the value of l/a having been fixed at 0.45 (see Fig. 16).

These results are, of course, merely illustrative, but they demonstrate that RNA should be a beautiful "laboratory system" for the study of gels with anomalous elastic properties. As Fig. 15 shows, one can tune the control parameter ν_2 by adjusting the extension, which can, in turn, be achieved via changes in the osmotic swelling pressure of the system. The second key parameter, the l/a ratio, can be "programmed" into the RNA molecules by alternating random sequences with complementary homopolymer sequences of prescribed length (e.g., strings of C monomers alternating with strings of G monomers). The most challenging feature would be to generate alignment between the rods to produce a nematic elastomer. Curiously, large SS RNA molecule n the form of 1400 base-long viral genomes are found to be quite anisotropic according to low angle x-ray diffraction and light scattering studies [26]. This suggests that large SS RNA molecules may be *naturally* anisotropic. Assembly of the gel under mild shear flow may enhance this natural anisotropy.



FIG. 15. The quantity ν_2 as defined in Eq. (2.5) and as derived from the fitting curve in Fig. 13.

In summary, RNA gels are expected to be rich laboratory systems for the study of fundamental elasticity.

A. Effects of disorder

The systems that we have considered in this paper are anisotropic and ordered. The effects of structural disorder, likely to occur in systems such as RNA gels, constitute an important issue. Through their secondary structure, RNA molecules indeed are structurally heterogeneous. Quenched disorder is known to have nontrivial effects on the elastic properties of conventional isotropic gels, such as the "butterfly effect" in small-angle neutron scattering [27]. The model explored here treats large RNA molecules as a linked network of orientationally ordered rods. Experiments on nematogenic gels of this sort, originally cross-linked in the isotropic phase, demonstrate that such gels, in fact, lose their long-range orientational order by the action of the quenched random stresses; the sample breaks up into a polydomain structure. Under an applied stress, a structural change can take place towards a more ordered monodomain structure, the *PM* transition [28]. The theoretical study by Uchida [29] on the effects of quenched disorder on nematic gels, in terms of soft modes, confirms that the disorder destroys long-range orientational order and that external stress can partially restore orientational order. Quenched disorder thus appears to play an even more important role for nematogenic gels.

The focus of this paper is on the Poisson ratio. Assume that, in the absence of external stress, disorder suppresses the long-range orientational order of a large RNA molecule. The structure breaks up into a polydomain sample. One can still apply the results obtained here to individual domains. If



FIG. 14. The quantity ν_1 as defined in Eq. (2.5) and as derived from the fitting curve in Fig. 13.



a

(μm)

FIG. 16. The Poisson ratio μ derived from the parameters ν_1 and ν_2 displayed in Figs. 14 and 15. The ratio l/a is fixed at the value 0.45. Note that the parameter *a* on the horizontal axis of this plot refers to the distance between rod centers and does not coincide with the extension shown in Figs. 14 and 15.

these domains have a negative Poisson ratio, then the aggregate of elastically coupled domains is expected to have a negative Poisson ratio as well. We believe, but have not proven, that a simple orientational average over different domain orientations should constitute a reasonable approach to the inclusion of the effects of disorder on Poisson's ratio. It is obvious, however, that in the presence of external stresses, the Poisson ratio will have a strong "signature" at the *PM* transition. The description of that signature would require a full treatment of structural disorder, for example using the method of Uchida [29].

B. Implications for viral assembly

It is interesting to speculate about possible consequences of a negative Poisson ratio for RNA gels. A curious problem of viral assembly is the large discrepancy between the density of the genome in solution and the same genome encapsidated by the virus's protein shell. Encapsidation proceeds by self-assembly: a solution of capsid proteins and viral RNA molecules will spontaneously assemble to form infectious viruses under physiological conditions. The assembly is driven by generic electrostatic affinity between the proteins and the RNA molecules, though specific interactions are required to initiate assembly, see, for instance, [30]. The scenario for the encapsidation of single-stranded RNA genomes into spherical viruses is not fully understood, but Fig. 17 indicates a likely scenario. A partially condensed RNA molecule forms a condensation surface for capsid proteins of opposite (i.e., positive) charge, and a curved shell starts to form. The challenge for the assembly process is how compaction of the genome can be achieved. If the RNA material had a positive Poisson ratio, then compaction of the genome at the nucleoprotein interface by the electrostatic affinity would produce swelling of the remainder of the molecule that is not in close contact with the growing shell. It would



FIG. 17. The first steps in the assembly of a spherical RNA virus. In Fig. 16(a), the capsomeres are beginning to come together. They attach to the portion of the genome in immediate proximity to them. Figure 16(b) shows what happens to the RNA complex as the capsomeres assemble and the RNA is compressed horizontally if the elastic response of the complex is characterized by a positive Poisson ratio. Figure 16(c) shows what happens if the RNA complex has a negative Poisson ratio.

seem that completion of assembly is not possible. A negative Poisson ratio, on the other hand, would lead to collapse of the RNA material into the shell. It would be fascinating if model systems could be developed with, for instance, RNA gels in contact with a positively charged substrate to verify this scenario. We conclude that RNA gels with negative Poisson ratios would be well adapted for easy encapsidation.

ACKNOWLEDGMENT

The authors gratefully acknowledge support of the National Science Foundation through NSF Grant No. DMR 0404507.

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